

Enantioselective total synthesis of (–)-epoxyquinols A and B. Novel, convenient access to chiral epoxyquinone building blocks through enzymatic desymmetrization

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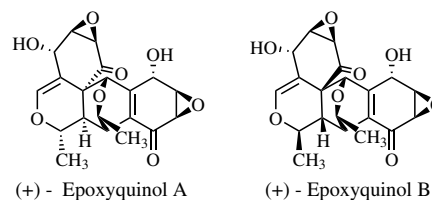
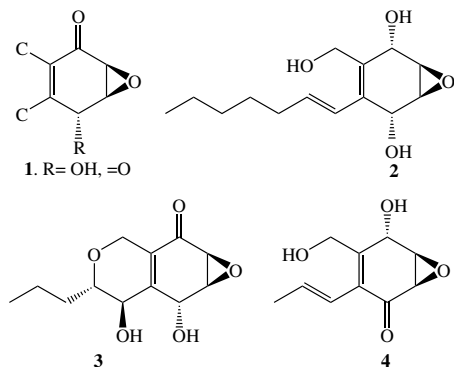
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Abstract—Following our recent total synthesis of the biologically potent natural products epoxyquinols A and B in racemic form, we have now accomplished the total synthesis of the (–)-epoxyquinols A and B, anti-podes of the angiogenesis inhibiting natural products, through a protocol that involves enzymatic desymmetrization of a versatile epoxyquinone derivative, readily available from the Diels–Alder adduct of cyclopentadiene and *p*-benzoquinone.

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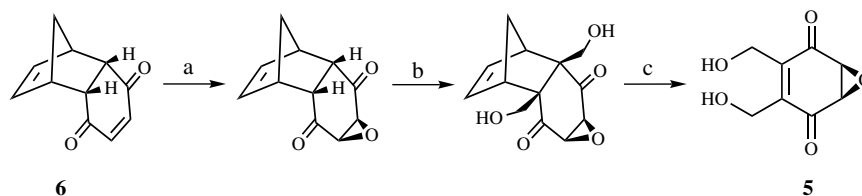
Several polyketide derived natural products, bearing the epoxyquinone moiety **1** as the core structure, have been encountered in nature from diverse sources in recent years.¹ Typical examples of such natural products are eupenoxide **2**,^{1a} cycloepoxydon **3**^{1b} and ECH **4**^{1c} among others, and many of them exhibit promising, wide ranging biological activity profiles. Quite naturally, epoxyquinone natural products have evoked considerable attention from the synthetic community over the past few years.² In our view, central to the synthetic approach to these natural products is a short, viable access route to the core structure **1** with adequately functionalized side arms. In this context, we have

recently disclosed a convenient three step access to the epoxyquinone building block **5** (cf. **1**)³ from the readily available Diels–Alder adduct **6** of cyclopentadiene and *p*-benzoquinone, Scheme 1. Recognizing the inherent versatility of **5** in the synthesis of epoxyquinone natural products, it was considered imperative to access its derivatives in enantiomerically pure form to harness fully its potential. The symmetrical nature of **5** lends itself amenable to the enzyme mediated ‘*meso* trick’. Indeed, the enzymatic desymmetrization of **5** has now been achieved and in this letter, we demonstrate the preparation of enantiomerically enriched derivatives of **5** and their utility in the total synthesis of (–)-epoxyquinols A and B.⁴

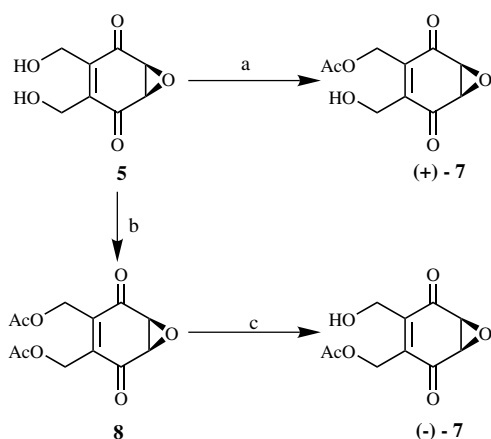


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After considerable experimentation with several lipases and reaction conditions, we were able to devise a simple, reproducible protocol for the enzymatic desymmetrization of **5** through transesterification, in a biphasic medium employing lipase PS 30 (Amano) immobilized on Celite, to furnish monoacetate (+)-**7** (~99% ee),



Scheme 1. Reagents and conditions: (a) 30% H₂O₂, 10% Na₂CO₃, acetone, 0 °C, 95%; (b) DBU, 40% formalin, THF, 92%; (c) diphenylether, 240 °C, 90%.



Scheme 2. Reagents and conditions: (a) Lipase PS 30 (Amano), vinyl acetate, *t*-butylmethylether, 0 °C, 6 h, 82%; (b) Ac₂O, pyridine, DMAP, DCM, 0 °C, 75%; (c) *C. rugosa* lipase, distd water, phosphate buffer, rt 86%.

Scheme 2.⁵ The absolute configuration of (+)-7 followed from its conversion to the natural products (–)-cycloepoxydon²ⁱ and (–)-epoxyquinols A and B (vide infra). Concurrently, *meso*-diol **5** was converted to the diacetate **8** and desymmetrization employing enzymatic hydrolysis with the same lipase PS 30 (Amano) gave (–)-7 (~86% ee) with modest enantioselectivity, Scheme 2. However, with the lipase from *Candida rugosa* (–)-7 was obtained from **8** in high enantiomeric purity (~99% ee).⁵ The versatile building block **7** thus became readily accessible in both its enantiomerically enriched forms through enantiodivergency based protocols. We demonstrate here the utility of one of the enantiomers (+)-7 towards the enantioselective synthesis of epoxyquinone natural products (–)-epoxyquinol A **9** and (–)-epoxyquinol B **10**.

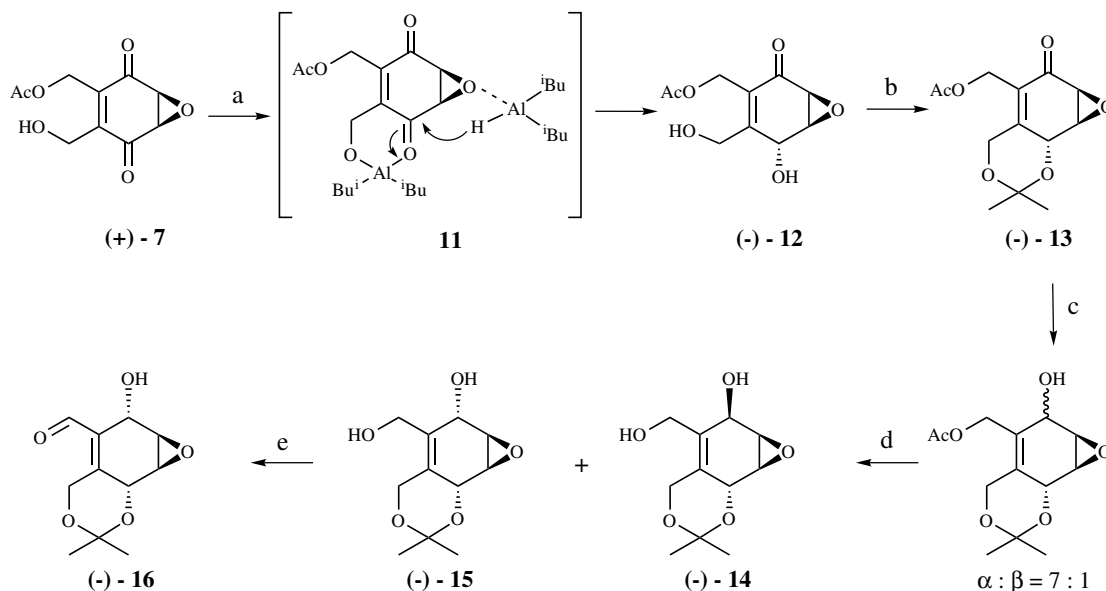
Angiogenesis inhibition is emerging as a very promising protocol with therapeutic potential for a variety of disorders ranging from rheumatoid arthritis to cancer⁶ and several natural products are being explored as leads for clinical development.⁷ In this context, the report of the isolation of two pentaketide derived dimers (+)-epoxyquinol A and (+)-epoxyquinol B from a soil based unknown fungus in 2002, by Japanese scientists, has drawn the immediate attention of synthetic chemists as these unusual natural products exhibited impressive inhibition of angiogenesis.⁴ In a short span of one year, three total syntheses of these natural products have

appeared in the literature.^{3,8} Two of these synthetic accomplishments by the groups of Hayashi^{8a} and Porco^{8b} are enantioselective and have led to the natural enantiomers (+)-epoxyquinol A and (+)-epoxyquinol B. On the other hand, our group has reported a synthesis of racemic epoxyquinols A and B.³ In addition, Hayashi and co-workers have delineated a practical approach to both the enantiomers of epoxyquinols A and B and also disclosed the preparation of some related model compounds.^{1c,9}

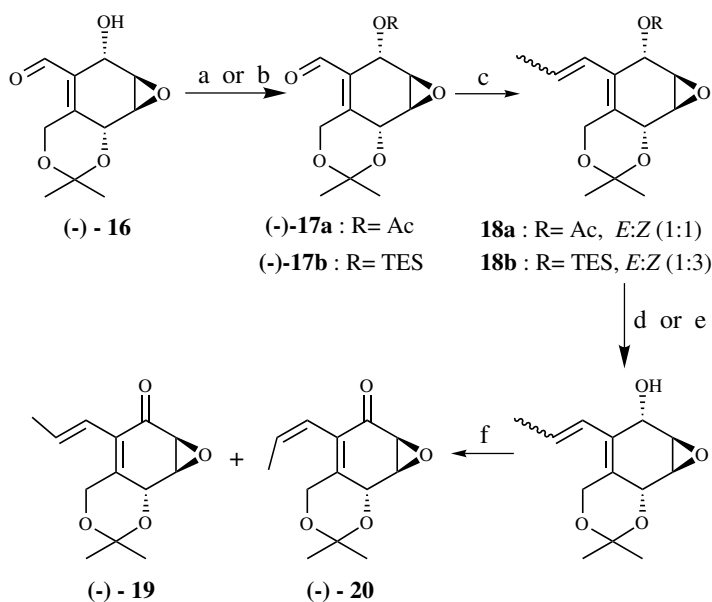
Our synthesis of (–)-9 and (–)-10, patterned along a biomimetic route, emanated from (+)-7 and a modified and improved variant of our previously described synthesis of racemic **9** and **10** was executed.³ DIBAL-H reduction of (+)-7 was regio- and stereoselective¹⁰ and directed by the primary hydroxyl group and epoxide oxygen and furnished diol (–)-12 through the intermediacy of the aluminium chelate **11**, Scheme 3.¹¹ The diol moiety in **12** was protected as the acetonide (–)-13. Reduction of the carbonyl group in **13** with Luche reagent¹² was moderately stereoselective, directed through the coordination of the ceric ion with the epoxy oxygen, to furnish a 7:1 diastereomeric mixture, which on further acetate hydrolysis led to (–)-14 and (–)-15. The relative stereochemical issue was settled through the X-ray crystal structure determination¹³ of the minor diastereomer (–)-14, Scheme 3.¹¹ The primary hydroxyl group in the major epimer (–)-15 was chemoselectively oxidized with TEMPO¹⁴ to furnish the aldehyde (–)-16.¹¹

Although hydroxy-aldehyde (–)-16 was ready for the installation of the requisite side chain, we found that without the protection of the hydroxyl group, Wittig olefination was capricious. Protection of the hydroxyl group as acetate **17a** and Wittig olefination furnished a 1:1 mixture of *E* and *Z* isomers **18a** in moderate yield. Protection of the hydroxyl group in **16** as TES ether **17b** and Wittig olefination gave **18b** in improved yield and an altered *E:Z* ratio of 1:3, Scheme 4. For further efforts, both **18a** and **18b** were serviceable and after hydroxyl group deprotection and oxidation, diastereomeric (*E*)-dienone (–)-19 and (*Z*)-dienone (–)-20 were readily separated and fully characterized, Scheme 4.¹¹

The acetonide deprotection in (–)-19 and (–)-20 led to the diols (–)-21 and (–)-22, respectively, Scheme 5.¹¹ TEMPO mediated¹⁴ chemoselective oxidation of (–)-21 and (–)-22 furnished aldehydes **23** and **24**, respectively, to set up the pericyclic cascade of 6π electrocyclozation



Scheme 3. Reagents and conditions: (a) DIBAL-H, THF, $-78\text{ }^{\circ}\text{C}$, 74%; (b) DMP, PPTS, acetone, rt 89%; (c) NaBH_4 , $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$, MeOH, $0\text{ }^{\circ}\text{C}$, 86%; (d) K_2CO_3 , MeOH, $0\text{ }^{\circ}\text{C}$, (-)-14 (11%) and (-)-15 (74%); (e) TEMPO, O_2 , CuCl, DMF, 76%.

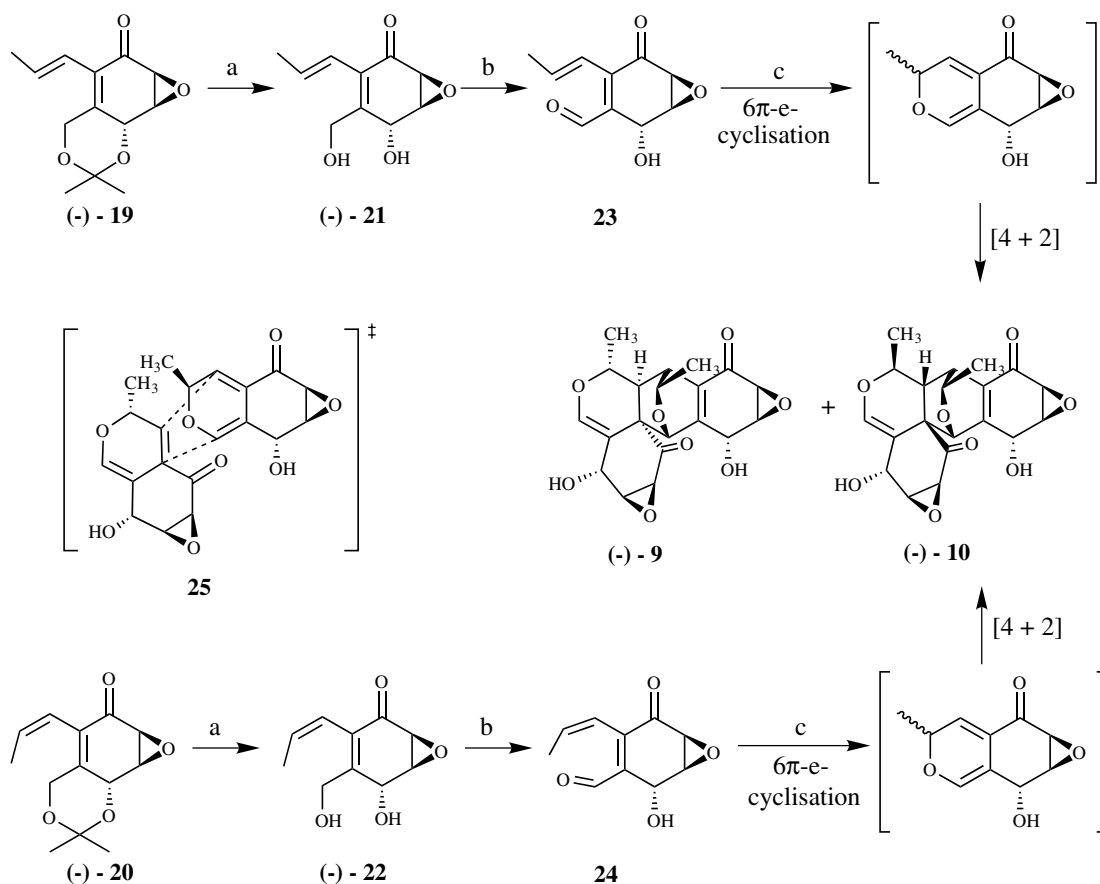


Scheme 4. Reagents and conditions: (a) Ac_2O , pyridine, DMAP, DCM, $0\text{ }^{\circ}\text{C}$, 87%; (b) TESCl, imidazole, DMAP, DCM, $0\text{ }^{\circ}\text{C}$, 70%; (c) $\text{C}_2\text{H}_5\text{PPh}_3\text{Br}$, $n\text{-BuLi}$, THF, $0\text{ }^{\circ}\text{C}$ (52% for **18a**, 76% for **18b**); (d) K_2CO_3 , MeOH, $0\text{ }^{\circ}\text{C}$, 90%; (e) HF·py, THF, $0\text{ }^{\circ}\text{C}$, 78%; (f) PDC, DCM, $0\text{ }^{\circ}\text{C}$, 81%.

and [4+2]-cycloaddition as depicted in Scheme 5. When aldehydes **23** and **24** were left aside neat at ambient temperature, the desired cyclization–cycloaddition occurred readily. In the (*E*)-series (–)-epoxyquinols (–)-**9** and (–)-**10** were obtained in a ratio of 3.5:1 from **21**, Scheme 5. On the other hand, in the (*Z*)-series (–)-epoxyquinols (–)-**9** and (–)-**10** were realized in a ratio of 4:1 in comparable yield.¹⁵ While an hetero-[4+2]cycloaddition through an *endo*-transition state **25** accounts for the formation of (–)-**9**, an analogous homo-[4+2] cycloaddition through an *exo*-transition state would result in the formation of (–)-**10**. It is interesting to note

that both the diastereomers (–)-**21** and (–)-**22** are serviceable and undergo the projected electrocyclization–cycloaddition cascade process with equal felicity and efficiency to furnish the epoxyquinol natural products.

In short, we have outlined a simple, convenient access to enantiomerically enriched, versatile building blocks (+)-**7** and (–)-**7** from the readily available Diels–Alder adduct of cyclopentadiene and *p*-benzoquinone. The former has been elaborated to (–)-epoxyquinols **A 9** and **B 10**, following a biomimetic pathway.



Scheme 5. Reagents and conditions: (a) Amberlyst 15, MeOH, rt (79% for **21**, 73% for **22**); (b) TEMPO, O₂, CuCl, DMF, rt; (c) neat, ~30 °C, 8 h, (-)-**9** (48%) and (-)-**10** (18%).

Acknowledgements

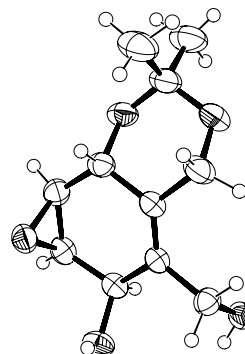
K.I. thanks CSIR, India for the award of a research fellowship. This work was supported by the Chemical Biology Unit of JNCASR at the Indian Institute of Science, Bangalore. The lipase used in this study was a gift from Dr. Y. Hirose of Amano Pharmaceutical Co. Ltd, Japan.

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- Enantiomeric excess (ee) was determined through ¹H NMR analyses (integration of the acetate methyl groups) after the addition of chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+)-camphorato]europium (III). *Procedure for enzymatic transesterification*: A mixture of *meso*-diol **5** (2 g, 10.9 mmol), vinyl acetate (4.09 mL, 43.6 mmol) and Amano lipase PS-30 immobilized on Celite (2 g) in *t*-butylmethylether was stirred for 6 h at 0 °C. The reaction mixture was filtered through a pad of Celite and the filtrate was concentrated. The crude product was subjected to column chromatography on silica gel and elution with hexane/ethyl acetate (2:1) furnished (+)-**7** (2.0 g, 82%) adjudged pure spectroscopically, as a colourless oil; [α]_D²⁴ +11.5 (*c* 3.2, CHCl₃). *Procedure for enzymatic hydrolysis*: A mixture of *meso*-diacetate **8** (50 mg, 0.187 mmol) and lipase from *C. rugosa* (Sigma-Aldrich) in distilled water (1.5 mL) was stirred at room temperature for 40 h. The reaction mixture was kept neutral by occasional addition of phosphate buffer (pH 7). After the usual work-up involving phosphate extraction with ethyl acetate, the crude product was subjected to column chromatography on silica gel to furnish (-)-**7** (28 mg,

- 86% based on recovered starting material) as a colourless oil; $[\alpha]_{\text{D}}^{24} -11.6$ (c 1.5, CHCl_3).
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 - X-ray data for (–)-**14**: X-ray data were collected at 293 K on a BRUKER SMART APEX CCD diffractometer with graphite monochromated $\text{Mo K}\alpha$ radiation ($\lambda = 0.7107 \text{ \AA}$). Structure was solved by direct methods (SIR92). Refinement was done by full-matrix least-squares procedures on F^2 using SHELXL-97. The nonhydrogen atoms were refined anisotropically whereas hydrogen atoms were refined isotropically. $\text{C}_{11}\text{H}_{16}\text{O}_5$, MW = 228.24, colourless crystal, crystal system: monoclinic, space group: $C2/c$, cell parameters: $a = 37.508$ (11) Å ,

$b = 6.415$ (2) Å , $c = 9.288$ (2) Å , $\beta = 92.690$ (5), $V = 2245.2 \text{ \AA}^3$, $Z = 8$, $D_c = 1.35 \text{ g cm}^{-3}$, $F(000) = 976.0$, $\mu = 0.11 \text{ mm}^{-1}$. Total number of l.s. parameters = 209, $R1 = 0.0436$ for 2028 $F_o > 4\sigma(F_o)$ and 0.0534 for all 2452 data. $wR2 = 0.1090$, GOF = 1.011, restrained GOF = 1.011 for all data (CCDC 234194). ORTEP diagram is shown below



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- Specific rotation for epoxyquinol A: $[\alpha]_{\text{D}} (-)-58^\circ$ (c 0.43, MeOH), [lit. $[\alpha]_{\text{D}} (+)-61^\circ$ (c 0.15, MeOH)]^{4a} and for epoxyquinol B: $[\alpha]_{\text{D}} (-)-146^\circ$ (c 0.52, MeOH), [lit. $[\alpha]_{\text{D}} (+)-153^\circ$ (c 0.32, MeOH)].^{4b}